

# EmMa.Test

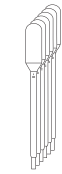
## Major + Minor XM Feline

QR CODE : Movie Procedure



### PROCEDURE FOR FELINE MAJOR + MINOR CROSSMATCH

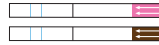
Material provided



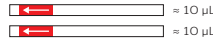
6 pipettes



2 large tubes  
for plasma



2 EmMa membranes



2 blood collector strips



1 blue top  
buffer



2 wash  
buffers



2 small tubes



1 yellow  
top buffer

**MAJOR** = Donor's Red Blood Cells (RBCs) + Recipient's Plasma

**MINOR** = Recipient's RBCs + Donor's Plasma

### N°1 : PREPARATION OF BLOOD SAMPLES AND TRAY

1

Centrifuge **DONOR** and **RECIPIENT** blood tubes **5 minutes at 1000g** to obtain packed red cells (pRBCs) and plasma<sup>(1)</sup>.

**DONOR**

Whole blood

- EDTA
- CPD
- ACD

Do not use Heparin



← Plasma

← pRBCs

**RECIPIENT**

Whole blood

- EDTA
- CPD
- ACD

Do not use Heparin



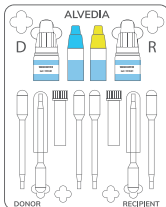
← Plasma

← pRBCs

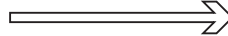
<sup>(1)</sup> or serum if using a dry tube

2

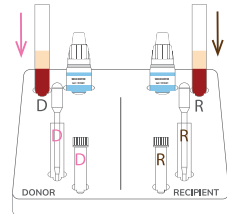
**KEEP**  
the material  
provided  
in the tray



Place vertically :  
the blood tubes, the wash buffers,  
the large and small tubes  
as follow



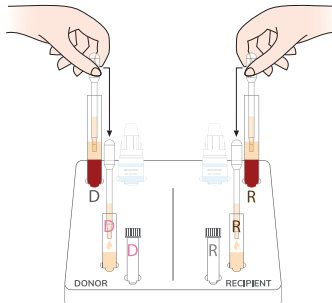
Write down «D» and «R»  
on the large and small tubes  
as follow



3

Take the pipette  
in the **D** large tube and transfer  
**the entire plasma only**  
from the donor's blood tube  
to the **D** large tube.

Discard the pipette.



Take the pipette  
in the **R** large tube and transfer  
**the entire plasma only**  
from the recipient's blood tube  
to the **R** large tube.

Discard the pipette.

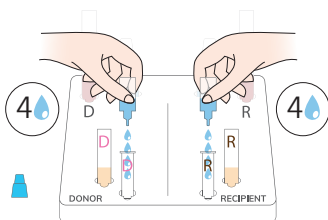


**KEEP YOUR ORIGINAL DONOR'S AND RECIPIENT'S BLOOD TUBES CONTAINING PRBCS**

## N°2 : PREPARATION OF MAJOR AND MINOR XM

### 1 Take the only one blue top buffer for both small tubes

Add 4 drops of the blue top buffer into the **D** small tube and 4 drops in the **R** small tube



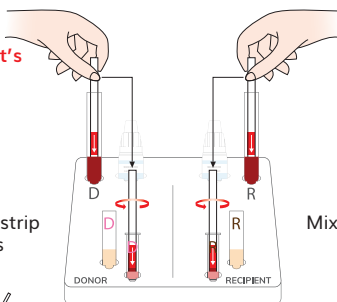
### 2 Vortex briefly donor's and recipient's blood tubes containing pRBCs

Dip the extremity of the blood collector strip into pRBCs DONOR's blood tube (10µL of pRBCs)

Mix the extremity of the blood collector strip in the **D** small tube during 7 seconds



Discard blood collector strip



Dip the extremity of the blood collector strip into pRBCs RECIPIENT's blood tube (10µL of pRBCs)

Mix the extremity of the blood collector strip in the **R** small tube during 7 seconds



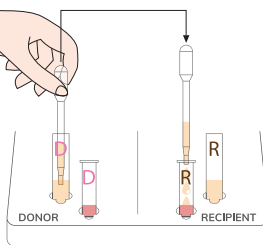
Discard blood collector strip



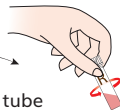
### 3 Take a new pipette and collect the DONOR's plasma from the **D** large tube and add 3 drops of plasma in the **R** small tube containing pRBCs suspension.



Discard the pipette



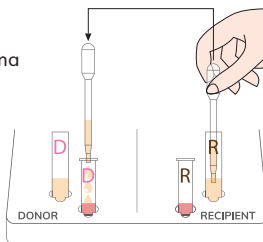
Close the **R** small tube and Mix gently



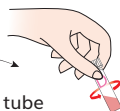
Take a new pipette and collect the RECIPIENT's plasma from the **R** large tube and add 3 drops of plasma in the **D** small tube containing pRBCs suspension.



Discard the pipette



Close the **D** small tube and Mix gently



**DISCARD BOTH **D** AND **R** LARGE TUBES CONTAINING THE REMAINING PLASMA.**

## N°3 : INCUBATION

Incubate both **D** and **R** tubes at room temperature during 10 minutes



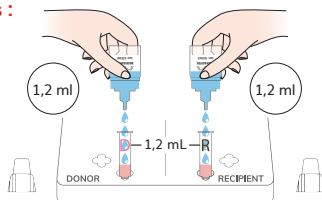
## N°4 : WASHING PROCEDURE

### FIRST WASH

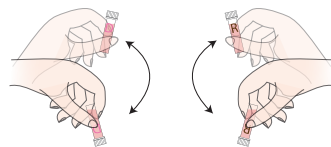
1

Take the wash buffers :  
1 for each small tube.

Fill both small tubes  
up to 1,2 ml  
of wash buffer



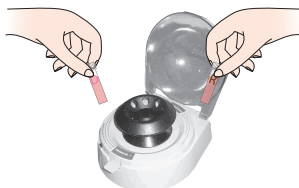
2



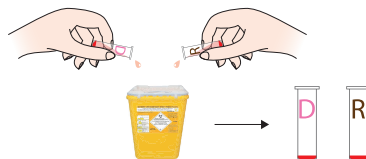
Mix both suspensions 3 times minimum

3

Centrifuge  
both tubes



4

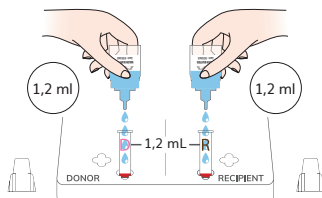


Discard the supernatant only :  
the RBCs pellet will stay at the bottom

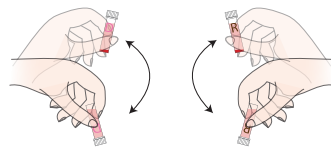
### SECOND WASH

5

Fill both small tubes  
up to 1,2 ml  
of wash buffer



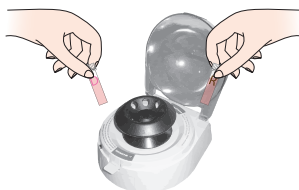
6



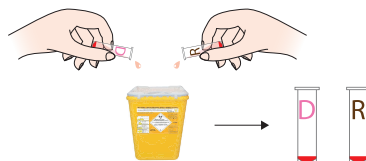
Resuspend completely the pellet  
by mixing both suspensions several times

7

Centrifuge  
both tubes



8

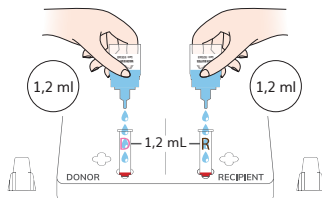


Discard the supernatant only :  
the RBCs pellet will stay at the bottom

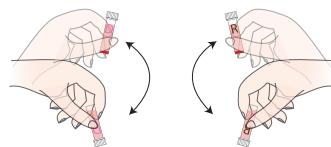
### THIRD WASH (PROCEDURE TO AVOID DILUTION BEFORE TESTING)

9

Fill both small tubes  
up to 1,2 ml  
of wash buffer



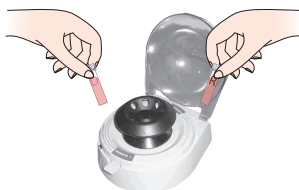
10



Resuspend completely the pellet  
by mixing both suspensions several times

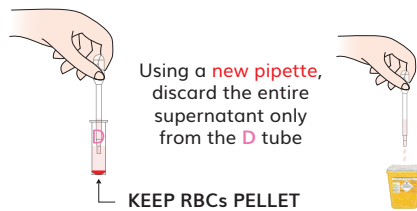
11

Centrifuge  
both tubes



12

A



Using a **new pipette**,  
discard the entire  
supernatant only  
from the **D** tube

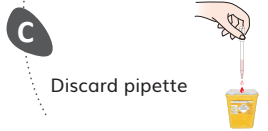
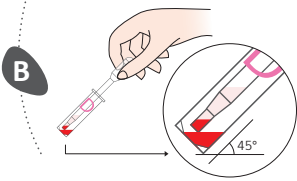
KEEP RBCs PELLET

Remove the residual supernatant until reaching few RBCs

Discard the remaining supernatant with few RBCs

Perform the steps 12.A .B .C for the R tube

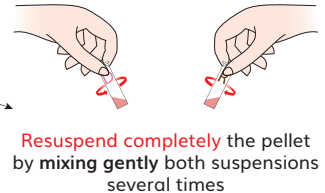
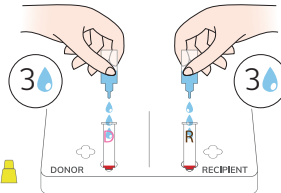
Washed RBCs pellets ready for XM test procedures



## N°5 : XM TEST PROCEDURE

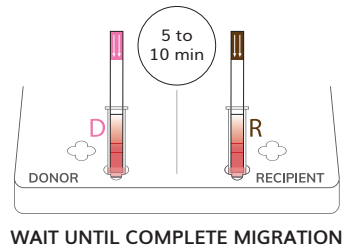
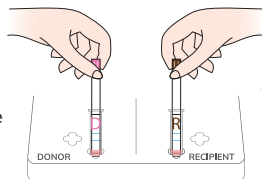
1 Take the only one yellow top buffer for both tubes

Add 3 drops of the yellow top buffer in the **D** tube and 3 drops in the **R** tube.

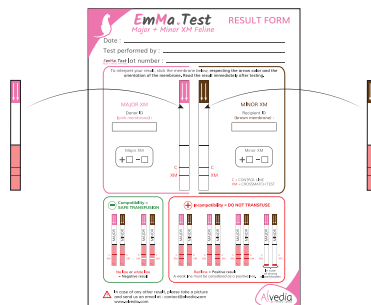


2 Insert the **PINK** EmMa membrane in the **D** tube

Insert the **BROWN** EmMa membrane in the **R** tube



3 Read the XM results at the end of the migration by sticking both membranes on the result form respecting the arrow colors



## SCIENTIFIC ADVISES

It is **MANDATORY** to blood type the donor and the recipient before making a Feline XM Test. Always transfuse **COMPATIBLE** blood.

Be careful, low titer and/or low affinity alloantibodies can be eluted during washing step procedures. This can affect the sensitivity of the XM test (e.g. low affinity/titer of anti-B in A blood group cat's plasma)

Usually, these alloantibodies cannot induce severe or mild hemolytic transfusion reaction.

Troubleshooting  
Please contact the Scientific Service Laboratory  
contact@alvedia.com  
+33(0)478 380 239

**Alvedia**  
Allice Veterinary Diagnostic